

## THE YEAST *YBR235W* GENE ENCODES A HOMOLOG OF THE MAMMALIAN ELECTRONEUTRAL $\text{Na}^+(\text{K}^+)\text{-Cl}^-$ COTRANSPORTER FAMILY

Bruno André <sup>\*,†</sup> and Bart Scherens <sup>#</sup>

\* Laboratoire de Physiologie Cellulaire et de Génétique des Levures  
Université Libre de Bruxelles, Campus Plaine CP 244  
Bld du Triomphe B-1050 Brussels, Belgium

# Research Institute of the CERIA-COOVI, Vlaams Interuniversitair  
Instituut voor Biotechnologie, Departement Microbiologie,  
Av. E. Gryson 1, B-1070 Brussels, Belgium

Received October 17, 1995

---

**Summary :** The *YBR235w* gene of the yeast *Saccharomyces cerevisiae* was found during sequencing of chromosome II. Here, we show that the 1120 aa protein (Ybr235p) encoded by this gene shares strong sequence similarity with the highly related electroneutral  $\text{Na}^+\text{-Cl}^-$  and  $\text{Na}^+\text{-K}^+\text{-Cl}^-$  cotransporters of animal cells. We hypothesize that this yeast protein also mediates active uptake of  $\text{Cl}^-$  into the cell. © 1995 Academic Press, Inc.

---

### Materials and Methods

To establish a preliminary classification of membrane transport proteins in *S. cerevisiae* (1), release 4.1 of the YPD database containing 4305 proteins (all Genbank *S. cerevisiae* sequences through July 7) (2) was screened for proteins having at least 4 predicted transmembrane domains (3;  $H_{19}$  method). Of the 301 proteins thus retrieved, those of known function uninvolved in transport were discarded. The remaining ones (most of which were discovered by genome sequencing) were compared with each other and used in protein similarity searches performed by means of the BLAST algorithm (4).

### Results and Discussion

Active transport of  $\text{Cl}^-$  ions into many epithelial and non-epithelial animal cells is ensured by electrically silent  $\text{Na}^+\text{-Cl}^-$  symporters, the driving force for  $\text{Cl}^-$  entry being provided by the energetically favourable uptake of  $\text{Na}^+$  down its electrochemical gradient (maintained by the  $[\text{Na}^+:\text{K}^+]$  ATPase) (5-6). The  $\text{Na}^+\text{-Cl}^-$  cotransporters can be grouped according to their sensitivities to inhibitors and their requirements for  $\text{K}^+$ . Thus, some mediate  $\text{Na}^+\text{-Cl}^-$

---

<sup>†</sup> To whom correspondence should be addressed. Fax: 32-2-6505421; Email: bran@ulb.ac.be.

cotransport tightly coupled to that of  $K^+$  with a stoichiometry of  $1Na^+-1K^+-2Cl^-$ . These  $Na^+-K^+-Cl^-$  cotransporters are sensitive to bumetanide and have been found in many different cell types. For instance, in the thick ascending limb of the loop of Henle in the kidney, epithelial cells reabsorb  $NaCl$  in excess water through an apical  $Na^+-K^+-2Cl^-$  cotransporter. Epithelial salt secretion involves a basolateral  $Na^+-K^+-2Cl^-$  cotransporter that is considered to provide the main entry pathway for  $Cl^-$  and  $Na^+$ . Other electroneutral  $Na^+-Cl^-$  cotransporters are  $K^+$ -independent, insensitive to bumetanide but highly sensitive to the thiazides. These  $Na^+-Cl^-$  cotransporters have been identified in renal and non-renal epithelia. The genes encoding thiazide-sensitive  $Na^+-Cl^-$  and bumetanide-sensitive  $Na^+-K^+-Cl^-$  cotransporters have been recently isolated (5-6). The deduced amino acid sequences defined a new family of highly similar proteins presumably made of 12 membrane-spanning regions flanked by large cytoplasmic N- and C-termini. It was speculated that other cation- $Cl^-$  cotransporters, such as bumetanide-sensitive  $Na^+-Cl^-$  cotransporters or the  $K^+-Cl^-$  cotransporters detected in erythrocytes and epithelia, could also be protein members of this family (5).

In the course of a systematic computer-aided analysis of putative transport proteins in the yeast *Saccharomyces cerevisiae* (1), our attention was drawn by the *YBR235w* gene of chromosome II (7). This gene encodes a 1120 amino acid protein with 12 predicted transmembrane domains and sharing sequence homology with the  $Na^+-(K^+)-Cl^-$  cotransporter family described above. Figure 1 shows the sequence alignment of the yeast *YBR235w* product with the secretory  $Na^+-K^+-Cl^-$  cotransporter from shark rectal gland epithelia (sCCC1) (8), the mammalian absorptive  $Na^+-K^+-Cl^-$  cotransporter of basolateral kidney cells (rCCC2) (9-10) and the thiazide-sensitive  $Na^+-Cl^-$  cotransporter from the urinary bladder of the teleost *P. americanus* (tCCC3) (11). The highest degree of amino-acid identity is in the predicted transmembrane regions. The yeast Ybr235 protein and the vertebrate  $Na^+-(K^+)-Cl^-$  cotransporters share 30-32 % identical residues within this region. The predicted intracellular loop connecting predicted transmembrane segments 2 and 3 was previously noted as being extremely well conserved among members of the Na-K-Cl family; this region is as well conserved in yeast Ybr235p (Fig. 1). The C-terminal hydrophilic extremity of Ybr235p is much smaller compared with those of vertebrate  $Na^+-(K^+)-Cl^-$  cotransporters.

The high degree of similarity between Ybr235p and vertebrate  $Na^+-(K^+)-Cl^-$  cotransporters suggests that Ybr235p might also correspond with an active uptake system for  $Cl^-$  ion. This putative  $Cl^-$  transporter could function as a  $H^+-Cl^-$  symporter or a  $Na^+-Cl^-$  symporter. Although the vacuolar membrane of yeast cells has been shown to contain  $Cl^-$  transport systems contributing to the formation of a chemical  $H^+$  gradient across the vacuolar membrane (13), there is as yet no reported experimental evidence of the existence of an active  $Cl^-$  uptake system in the plasma membrane of yeast cells. Imaginably, such a  $Cl^-$  cotransporter might play a role in regulating the water content of yeast cells, since the role of  $Na^+-K^+-Cl^-$



6. Payne, J.A. and Forbush, B. (1995) *Cur. Biol.* 7, 493-503.
7. Feldmann, H., Aigle, M., Aljinovic, G., André, B et al. (1994) *EMBO. J.* 13, 5795-5809.
8. Xu, J.C., Lytle, C., Zhu, T.T., Payne, J.A., Benz, E., Jr., and Forbush, B. (1994) *Proc. Natl. Acad. Sci. U. S. A.* 91, 2201-2205.
9. Payne, J.A., and Forbush, B. (1994) *Proc. Natl. Acad. Sci. U. S. A.* 91, 4544-4548.
10. Gamba, G., Miyanoshita, A., Lombardi, M., Lytton, J., Lee, W.S., Hediger, M.A., and Hebert, S.C. (1994) *J. Biol. Chem.* 269, 17713-17722.
11. Gamba, G., Saltzberg, S.N., Lombardi, M., Miyanoshita, A., Lytton, J., Hediger, M.A., Brenner, B.M., and Hebert, S.C. (1993) *Proc. Natl. Acad. Sci. U. S. A.* 90, 2749-2753.
12. Versaw, W.K., and Metzenberg, R.L. (1995) *Proc. Natl. Acad. Sci. U. S. A.* 92, 3884-3887.
13. Wada, Y., Ohsumi, Y., and Anraku, Y. (1992) *Biochim. Biophys. Acta* 1101, 296-302.